Molecular Insights into the Development and Efficacy of Hepatitis Virus Vaccines

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Viral hepatitis remains a major public health challenge globally, especially hepatitis B and C. This puts millions of people at risk of various forms of chronic liver diseases, such as liver cirrhosis, hepatocellular carcinoma, liver failure, etc. Vaccines have been confirmed to be effective in reducing the burden of certain infectious agents, including many of the hepatitis viruses. This article evaluates the molecular mechanisms involved in the design of the available hepatitis viruses, their efficacy, and how to improve them. HAV vaccines are designed around the formalin-inactivated virus or genetically attenuated live viruses, and molecular design emphasizes robust immunogenicity and safety. Current HBV vaccines are primarily recombinant subunit vaccines using the HBV surface antigen (HBsAg) and are produced by expressing HBsAg in yeast or mammalian cell systems, eliciting strong immune responses. HCV lacks an approved vaccine despite significant research efforts.

Its high genetic variability and immune evasion mechanisms make it complex in the development of vaccines. However, most HCV vaccine candidates focus on conserved epitopes within the viral envelope glycoproteins E1 and E2, which are critical for viral entry. Currently, there are no standalone vaccines for Hepatitis although the infection can be prevented by hepatitis B immunization. Moreover, efforts to develop HDVspecific vaccines focus on the large HDV antigen (HDAg), which is essential for viral assembly. The first licensed HEV vaccine, Hecolin®, is a recombinant subunit vaccine based on the ORF2 protein of HEV genotype 1 and is produced in Escherichia coli, where the ORF2 protein self-assembles into VLPs, mimicking the native virus's immunogenicity. DNA recombinant technology, protein engineering, structural biology applications, the use of virus-like particles, mRNA nanoparticle-based platforms, etc, have all increased the efficacy of the available vaccines.

1.0. INTRODUCTION

Viral hepatitis is the inflammation of the liver following an infection by any of the major hepatitis viruses A, B, C, D, and E. Other viruses implicated include cytomegalo virus, Herpes simplex virus, Epstein Barr virus, Rubella virus, Adenovirus, Mumps virus, Yellow fever virus, etc. When the inflammation does not resolve in

six months, the name changes from acute to chronic hepatitis. Some of the common risk factors for viral hepatitis include intravenous drug unprotected sexual intercourse, food and water contamination, blood transfusion, being a healthcare worker, being born to a hepatitis B or C-infected mother, etc.

DOI: 10.21608/aeji.2025.376694.1468

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For instance, the Hepatitis B virus (HBV) is a major public health challenge globally, with more than 30% of the global population having serological evidence of ongoing or previous infection. Chronic HBV accounts significantly for the global mortality rate, with almost a billion individuals dying annually from HBV-related hepatic diseases [1].

Vaccines are effective means of preventing and controlling many communicable diseases. The World Health Organization (WHO)'s Immunization Agenda 2030 (IA2030) emphasizes the need for global strategies to ensure everyone gets vaccinated [2]. The IA2030 highlights the ongoing challenges of infectious diseases and focuses on approaches to promote development, manufacturing, uptake, to ensure global coverage, through the efforts of researchers, healthcare providers, manufacturers, policymakers, and governmental non-governmental organizations Research has shown that vaccines are effective in significantly reducing worldwide HBV infection prevalence, primarily via childhood HBV vaccination which reduced the prevalence from 4.9% in the prevaccination period to 0.9% in 2019 [1]. This review explores the molecular mechanisms involved in the design of hepatitis virus vaccines, evaluating their efficacy and potential areas of improvement. The overall goal is to contribute to the understanding of the design of more effective vaccines to reduce the burden of hepatitis globally.

2.0.Molecular Basis of Hepatitis Virus Infection and Pathogenesis

Hepatitis virus invades the host through diverse molecular interactions which are peculiar to each type of virus due to their unique molecular makeup. In Hepatitis A infection, transmission is by by fecal-oral route [2]. The HAV genome (7.5 kb) codes for only one polyprotein, which is usually cleaved into structural proteins (VP4, VP2, VP3, and VP1pX capsid antigens) and nonstructural proteins, such as 2B, 2C, 3A, 3B (the genome-linked protein, VPg), 3Cpro (a cysteine protease), and 3Dpol (RNA-dependent RNA polymerase). These proteins are integral to viral replication and assembly [3]. Through this means, HAV causes acute inflammation of the liver that generally resolves without leading to chronic conditions, followed by life-long immunity, which is unlike other hepatotropic viruses [4].

Hepatitis B virus transmission occurs through contact with contaminated blood or body fluids, as well as vertical transmission after birth [5] [6]. The viral genome has a partially double-stranded relaxed circular DNA (rcDNA), that replicates via an RNA intermediate [7]. It codes for key proteins including the viral DNA polymerase (reverse transcriptase), hepatitis B core antigen (HBcAg), HBsAg, hepatitis B e antigen (HBeAg), and the hepatitis B x protein (HBxAg). The survival of HBV is facilitated by the integration of viral DNA into the host genome as covalently closed circular DNA (cccDNA) during chronic infection, which also raises the risk of serious liver disorders such as cirrhosis, liver failure, and HCC [6] [8]. HBsAg envelops circulating HBV virions, sometimes referred to as "Dane particles," which are made up of three virus-coded surface proteins: large (L), middle (M), and small (S). Despite its poor affinity, the HBV virion attaches itself to the surface of hepatocytes by interacting with heparan sulfate proteoglycans (HSPGs). This is followed by interaction with the sodium taurocholate cotransporting polypeptide (NTCP) receptor, which has a high affinity for the pre-S1 domain of L-HBsAg. This contact could cause HBV to internalize into hepatic cells by endocytosis [6].

As a satellite virus, the Hepatitis D virus requires co-infection (simultaneous infection with HBV and HDV) or superinfection (HDV infection in a person with chronic HBV) for its propagation within humans, leading to the most severe form of viral hepatitis. The virus mediates its de novo entrance into hepatocytes and facilitates the release of its progeny by using HBsAg. Much like HBV, it is predominantly transmitted through parenteral exposure to contaminated body fluids, intravenous drug use, and, less commonly, through vertical transmission [9]. The HDV RNA genome is very heterogeneous, with eight different genotypes that differ in how the disease progresses and responds to therapy [10].

In hepatitis C infection, the viral lifecycle includes viral entrance, uncoating, and release of the genome into the cytoplasm, which is followed by RNA translation and replication, particle assembly, and egress [11]. Human

hepatocytes are the primary target cell of HCV infection in vivo, and the virus may effectively infect human hepatoma cell lines in vitro [12]. The attachment and internalization of HCV require four essential cellular components: occludin (OCLN), claudin-1 (CLDN), scavenger receptor class B type I (SR-BI), and CD81. Of these, the primary target for antibody-mediated neutralization is the interaction between E2 and CD81 [13]. Overall, Several host cell factors are required for virus translation, replication, and production. These include liver-specific microRNA-122 (miR-122) [14], autophagy proteins (i.e. Beclin-1, Atg4B, Atg5 and Atg12) [15], cyclophilin A (CypA) [16], lipid droplets (LDs), and the VLDL (very low-density lipoprotein) pathway, export utilizing apolipoprotein B and E (ApoB and ApoE) as cofactors [17] amongst others.

Hepatitis E Virus is the final major Hepatitis virus. Our knowledge of HEV's reproduction cycle, host cell contacts, and entrance pathways into hepatic cells has only recently improved, despite evidence that suggests outbreaks have occurred since ancient times. Unlike traditional enclosed viruses, the eHEV are infectious but do not have virus-encoded proteins on their surface [18]. Findings support the idea that the biogenesis of eHEV viral particles includes the highly selective sorting of viral capsids into multivesicular endosomes (MVEs) in a process that is dependent on endosomal sorting complexes required for transport (ESCRT), which is similar to how exosomes are produced [19]. HEV virions most likely are involved in non-specific interactions with clathrin during

their initial attachment and endocytosis. Like nHEV, they are trafficked to endolysosomes by the GTPase Ras-related protein Rab-5A (RAB5A) and RAB7A [20].

Cellular enzymes in lysosomes mediate the breakdown of eHEV membrane lipids [21]. The compartment and mechanism of eHEV capsid uncoating are still unclear. The endolysosome may serve just as a transitory space when the virus sheds its quasi-envelope. It is known that the endoplasmic reticulum-associated degradation pathway allows overexpressed ORF2 proteins to go into the cytoplasm [22].

3.0. Current Hepatitis Virus Vaccines: Molecular Design and Properties

Hepatitis virus vaccines utilize advanced molecular designs to prevent infections effectively. Hepatitis A vaccines are usually inactivated virus formulations inducing strong, long-lasting immunity [23], while hepatitis B vaccines are recombinant protein vaccines using the hepatitis B surface antigen (HBsAg) produced in yeast or mammalian cells, ensuring robust immunogenicity [24]. Furthermore, hepatitis C lacks an approved vaccine due to its genetic diversity, though novel candidates, like recombinant protein and mRNA-based vaccines [25] [26] [27]. Hepatitis D prevention relies on the hepatitis B vaccine, as HDV requires HBV for replication. Hepatitis E vaccines, such as the recombinant HEV239, target the viral capsid protein, providing protection primarily in endemic regions [28]. An overview of the hepatitis virus vaccine, its molecular design, and other properties are depicted in Table 1 below.

Table 1: Table highlighting the molecular designs and properties of the hepatitis virus vaccines.

S/N	Vaccine	Molecular Design	Other Properties	References
1.	Hepatitis A Virus Vaccine	- Primarily based on inactivated viral particles or live attenuated strains, with the molecular	 Stable under standard refrigeration conditions and often adjuvanted with aluminum hydroxide to enhance immune response. Induces protective 	[23] [29]

		design	antibodies (anti-HAV	
		emphasizin		
		robust	vaccinated individuals	
		immunogen		
		ty and safet	y.	
		- Designed		
		around the		
		formalin-		
		inactivated		
		virus or		
		genetically		
		attenuated 1	ive	
		viruses.		
		viruses.		
	II de Dar ar			
2.	Hepatitis B Virus Vaccine	- Primarily	- Elicits a robust	[30] [31]
		recombinan		[50] [51]
			1 '	
		subunit	seroprotection rates	
		vaccines us		
		the HBV	healthy individuals	
		surface	after the standard	
		antigen	three-dose regimen	
		(HBsAg)	- Adjuvants such as	
		- Produced by	y aluminum hydroxide	
		expressing	enhance the immune	
		HBsAg in	response.	
		yeast or		
		mammalian	ı	
		cell systems	s	
		ĺ		
3.	Hepatitis C Virus Vaccine	- Lacks an		
	•	approved	- The variability of	[27] [32]
		vaccine, but	t HCV strains remains a	
		candidates	barrier to universal	
		focus on	vaccine efficacy.	
		conserved epitopes	vaccine enreacy.	
		within the		
		viral envelo	ppe	
		glycoprotein E1 and E2,	ns	
		which are		
		critical for viral entry.		
		- Other		
		strategies		
		include recombinan	ıt	
		VLPs,		
		adenoviral vectors, and	1	
		mRNA	•	
1		vaccines		
		encoding		

		T		
		HCV antigens.		
4.	Hepatitis D Virus Vaccine			
		- There are no		
		standalone		
		vaccines for		
		Hepatitis D,		
		but it can be		
		prevented by		
		HBV		
		immunization.		
	** ***			[22]
5.	Hepatitis E Virus Vaccine			[33]
		- A recombinant subunit vaccine	 Safe, stable, and induces long-lasting immunity. 	
		based on the	long-lasting minitunity.	
		ORF2 protein		
		of HEV		
		genotype 1		
		- Produced in		
		Escherichia		
		coli, where the		
		ORF2 protein		
		self-assembles into VLPs,		
		mimicking the		
		native virus's		
		immunogenicit		
		у.		

4.0. Technological Advances in Hepatitis Vaccine Development

4.41. Role of DNA Recombinant Technology

The advent of recombinant DNA technology marked a transformative era in hepatitis vaccine development, enabling the creation of safer, more effective, and scalable solutions. Early breakthroughs in this field laid the groundwork for addressing the global burden of hepatitis B and E through innovative genetic engineering techniques. In the 1980s, recombinant DNA technology revolutionized hepatitis B vaccine production by enabling the synthesis of hepatitis B surface antigens (HBsAg) in yeast cells or other expression systems [34]. These vaccines offered significant advantages over traditional

plasma-derived options, demonstrating superior immunogenicity, enhanced safety, and scalability, which were crucial for widespread immunization efforts [34] [35]. Building on these early successes, researchers harnessed recombinant DNA technology to refine and optimize vaccine design. By constructing synthetic vaccines that predict viral epitopes, they improved the precision of immune responses, enhancing the vaccines' ability to target the virus effectively [36].

Similarly, the application of recombinant DNA technology to hepatitis E vaccine development has been instrumental in producing virus-like particles (VLPs). These VLPs mimic the virus's structure without carrying its genetic material, ensuring safety while achieving high efficacy

[37]. More recently, DNA-based vaccines have emerged as a promising platform. These vaccines introduce genetic material encoding viral antigens into host cells, eliciting robust immune responses. With advancements in vector design and delivery mechanisms, DNA vaccines are poised for further innovation, particularly in preclinical evaluations [38] [39]. The global rollout of recombinant hepatitis B vaccines has led to a dramatic reduction in the prevalence of hepatitis B, particularly among children under five [40] [41]. Compared to emerging platforms like mRNA vaccines, recombinant approaches remain highly profitable and manufacturingfriendly, particularly in addressing diseases like hepatitis [42]. Innovations in peptide-based vaccines further illustrate the utility of this technology. Bv deleting infection-causing components, peptide vaccines act as antigens to stimulate effective immune responses, enhancing both vaccine stability and specificity. The integration of nucleic acid-based platforms with recombinant DNA methodologies has opened new avenues for addressing viral infections. These combined approaches enhance vaccine efficacy, delivery, and adaptability, making them valuable tools for hepatitis management [43].

4.2. Role of Protein Engineering

A pivotal application of protein engineering is the development of recombinant hepatitis B vaccines, which incorporate engineered surface antigens to enhance immunogenicity. These antigens can then be precisely folded and modified, resulting in vaccines with improved efficacy and safety [44]. Key innovations include modifications to the pre-S region of the hepatitis B virus envelope protein, which significantly enhance its immunogenic potential. These engineered vaccines have shown superiority over traditional formulations in both preventing hepatitis B infection and treating chronic hepatitis B [45]. Furthermore, incorporating hepatitis C virus epitopes into engineered chimeric proteins has demonstrated promise for developing multivalent vaccines targeting both hepatitis B and C [46.[

Advances in expression systems such as Pichia pastoris yeast have further bolstered vaccine development. This system has been used to produce glycosylated recombinant proteins like CoreE1E2, which induce neutralizing antibodies

against the hepatitis C virus, highlighting the role of protein engineering in optimizing antigen production for stronger immune responses [47]. Additionally. computational tools immunoinformatics have been integrated into protein engineering to design multiepitope peptide vaccines. Techniques such as protein docking and molecular dynamics simulations allow for the identification and optimization of antigenic regions, expanding the scope of engineered proteins in vaccine design [48].

Innovative approaches such as engineering fusion proteins that combine immunogenic components from different hepatitis viruses further expand the potential for broader immune coverage. This strategy holds promise for the development of universal hepatitis vaccines [49]. Protein engineering and recombinant DNA technology are intricately linked. Recombinant DNA technology provides the foundational tools for protein engineering by enabling the cloning. expression, and genetic modification of genes encoding specific viral proteins. The synergy between these technologies is particularly evident in the engineering of pre-S regions of HBsAg, which have been modified using recombinant DNA technology to enhance their ability to elicit neutralizing antibodies [45]. recombinant DNA Similarly, techniques facilitate the production of fusion proteins that combine epitopes from multiple viruses, such as hepatitis B and C. These fusion proteins, optimized through protein engineering, offer a novel approach to creating multivalent vaccines [49]. In addition, recombinant DNA technology enables the synthesis of multi-epitope genes that serve as templates for protein engineering [48]. Their complementary roles enable the creation of safe, effective, and cutting-edge vaccines. This integration continues to expand the possibilities for combating hepatitis and other infectious diseases through advanced vaccine technology.

4.3. Use of Virus-like Particles (VLPs) in **Hepatitis Vaccine Design**

Virus-like particles (VLPs) are highly promising platforms in hepatitis vaccine design due to their ability to mimic the structural properties of natural viruses while remaining non-infectious. VLPs are self-assembled from viral structural proteins, enabling the preservation of native antigenic conformations, which is crucial for eliciting strong immune responses [50]. Their non-replicating nature ensures a high safety profile, making them ideal candidates for developing vaccines against hepatitis B, C, and E viruses [51]. In hepatitis B vaccine development, VLPs derived from the hepatitis B surface antigen (HBsAg) have become the cornerstone of preventive strategies. These VLPs not only elicit robust antibody responses but also maintain a high degree of purity and immunogenicity through advanced expression systems such as CHO cells [52].

For hepatitis C, VLPs incorporating oligomeric forms of envelope protein E2 (sE2) have shown potential in eliciting broadly neutralizing antibodies. These VLPs, produced using selfassembling platforms, represent a significant step toward effective HCV vaccine development, overcoming challenges associated traditional subunit vaccines [53]. Hepatitis E vaccines have also leveraged VLP technology. utilizing ORF2-derived particles that mimic the natural virus structure while ensuring safety by eliminating viral replication. These VLPs have demonstrated high immunogenicity, marking progress in addressing the public health burden of hepatitis E [51]. VLPs are further enhanced by their ability to incorporate multiple antigens or be chemically modified for improved stability and immune targeting. This versatility extends their application beyond hepatitis vaccines to other infectious diseases, underscoring their transformative potential in vaccine design [54].

4.4.Role of Structural Biology

The structural characterization of hepatitis B surface antigens (HBsAg) has also played a key role in developing vaccines with high purity and Structural biology, combined with recombinant DNA technology, has enabled biochemical characterization precise enhanced antigen folding. This synergy has led to the development of synthetic polypeptide vaccines with improved stability and efficacy. For example, mammalian expression systems have been utilized to produce structurally faithful hepatitis B surface antigens, critical for effective vaccine formulations [44] [55]. Recent breakthroughs in structural biology include the glyco-engineered production of HBV antigens to enhance immunogenicity. By leveraging Nglycosylation, researchers have improved antigen folding and immune recognition, contributing to more effective vaccine designs [56]. Additionally, computational tools such as immunoinformatics have enabled the identification of immunogenic epitopes on HCV proteins, facilitating the design of multi-epitope vaccines with enhanced immune responses [48.]

Advancements in 3D cell culture models have provided valuable platforms for studying hepatitis pathology and antigen behavior in environments that mimic in vivo conditions. These models have been pivotal in exploring antigen-antibody interactions and improving vaccine efficacy [57]. Structural analyses of HCV-neutralizing antibodies have further revealed vulnerabilities in the virus's envelope proteins, guiding the rational design of effective antigens [58]. Today, cryo-electron microscopy (cryo-EM) and X-ray crystallography are often integrated to maximize their complementary strengths. Crvo-EM offers detailed views of antigenic surfaces and antibody-binding sites, while crystallography confirms the atomic structures of antigen-antibody complexes. For example, this approach has been used to study HCV envelope glycoprotein E1E2, defining neutralizing epitopes critical for vaccine design [59]. Recent innovations combine cryo-EM with machine learning to map epitopes and optimize antigens, advancing structure-based vaccine development [60]. Together, these structural biology techniques continue to drive innovations in hepatitis vaccine development, offering comprehensive insights into viral antigens and immune interactions.

4.5. Role of mRNA and nanoparticle-based platforms in next-generation vaccines

mRNA and nanoparticle-based platforms are revolutionizing next-generation hepatitis vaccine development by combining the specificity and adaptability of mRNA technology with the protective and delivery capabilities nanoparticles. mRNA vaccines rely on lipid nanoparticles (LNPs) to shield the mRNA from degradation and enhance its uptake by antigenpresenting cells, which not only facilitates antigen expression but also acts as an adjuvant to boost immune responses [61]. The versatility of nanoparticles extends beyond LNPs, with gold nanoparticles offering chitosan and additional avenues for targeted delivery. For example, chitosan nanoparticles have been explored for nasal delivery of the hepatitis B surface antigen (HBsAg), effectively eliciting both mucosal and systemic immune responses [62]. Gold nanoparticles, meanwhile, serve as nano-adjuvants in hepatitis Ε vaccines. enhancing immune responses and stabilizing vaccine components [63].

Advancements in nucleoside-modified mRNA and LNP delivery systems have further enhanced the stability and immunogenicity of mRNA vaccines. These innovations became globally recognized during the COVID-19 pandemic, proving the platform's capability to produce highly effective vaccines rapidly [61]. This success has renewed interest in using mRNA technology to combat other infectious diseases, including hepatitis. Therapeutic mRNA vaccines for chronic hepatitis B virus (HBV) infections have shown promising preclinical results, demonstrating their ability to elicit robust memory T and B cell responses crucial for longterm immunity and virological suppression [64]. Furthermore, self-amplifying mRNA vaccines, which enhance antigen expression immunogenicity, are being explored for hepatitis vaccine development [65].

The integration of nanoparticle-based delivery systems has significantly enhanced the efficacy of mRNA vaccines against hepatitis. LNPs protect mRNA, facilitate targeted delivery to antigen-presenting cells, and address challenges such as hepatic immune evasion [61]. Emerging innovations, including circular mRNA and nonviral delivery platforms, are being developed to overcome remaining challenges, such as the need for repeat doses and potential reactogenicity [66].

5.0. Challenges and Future Directions in **Hepatitis Vaccine Development**

5.1. Challenges and Limitations in Hepatitis Vaccine Development

Current vaccines for hepatitis viruses, while effective in many cases, face significant challenges and limitations that hinder their universal success. For hepatitis B virus (HBV), vaccine efficacy is notably limited in certain populations. Up to 10% of healthy individuals and as many as 50% of immunocompromised individuals. such as those undergoing hemodialysis, fail to generate protective antibody levels following vaccination [67]. Furthermore, the emergence of HBV escape mutants has compromised vaccine effectiveness in regions with high endemicity [68]. These challenges underscore the need for improved formulations, booster strategies, and alternative approaches.

Logistical barriers also affect HBV vaccine distribution. Current vaccines require a robust cold chain for stability, a challenge in rural and resource-limited settings that restrict coverage. Moreover. while current HBV vaccines effectively prevent chronic infections in many cases, they fail to eradicate the virus in chronic carriers due to its ability to integrate into the host genome and evade immune responses. This limitation makes complete viral control challenging, even with therapeutic interventions

For hepatitis C virus (HCV), no effective vaccine currently exists, largely due to the virus's high genetic variability and rapid mutation rates, which enable immune evasion. The lack of optimal animal models and an incomplete understanding of protective immune responses further complicate vaccine development [70]. Additionally, the asymptomatic nature of many HCV infections and high vaccine development costs pose significant hurdles to awareness and prevention efforts [71]. While DNA-based vaccine approaches have shown promise, they often fail to elicit long-lasting immune responses, highlighting the need for further optimization [26].

HDV vaccine development, in particular, is fraught with unique challenges. The virus's dependence on HBV for replication complicates the identification of specific vaccine targets. Prophylactic **HBV** vaccination indirectly prevents HDV co-infection, but it does not address existing HDV infections in chronic HBV carriers, leaving a significant gap in prevention strategies [72]. Moreover, the lack of robust animal models and optimized preclinical platforms hampers HDV vaccine research [39]. Virus-like particle (VLP) vaccines have shown potential for HDV, but their development faces hurdles in ensuring immunogenicity, scalability, and the selection of effective adjuvants [73].

5.2. Addressing the Limitations and Hurdles

Efforts to address the challenges and limitations of hepatitis vaccine development and efficacy have focused on advancing vaccine technologies, optimizing adjuvants, and improving delivery systems. Targeted strategies, such as booster vaccinations and the incorporation of potent adjuvants, have shown promise in enhancing vaccine effectiveness. For instance, nanoparticle-based adjuvants significantly improve immunogenicity, addressing response rates in specific populations, including individuals with obesity or genetic polymorphisms [74].

The emergence of mRNA-based vaccines offers a transformative approach to combating the hepatitis B virus (HBV). Preclinical studies show that mRNA vaccines, formulated with lipid nanoparticles and administered in prime/boost regimens, elicit robust immune responses and improve antigen presentation [75]. Similarly, recombinant viral vectors and virus-like particles (VLPs) are being developed as platforms to enhance immune responses and tackle the genetic variability and immune evasion tactics of hepatitis viruses [27]. Improving vaccine accessibility and coverage is another critical focus area. Addressing vaccine hesitancy through sensitive communication culturally community engagement strategies is essential for increasing vaccination rates in underrepresented [76]. Additionally, expanding populations vaccination coverage at birth and targeting unvaccinated adults are vital steps toward meeting global hepatitis control targets [77].

To address the limitations in vaccine testing models, innovative outbred animal models, such as rodent hepacivirus systems, have been developed. These models provide valuable platforms for testing immunization strategies, particularly for hepatitis C virus (HCV) vaccines, and enhance their clinical relevance [78]. Genebased vaccines and physiologically based pharmacokinetic models are also being employed to optimize vaccine safety and efficacy, especially in vulnerable populations such as pregnant women, while adhering to stringent regulatory and safety protocols [79].

HCV vaccine development faces significant challenges due to the virus's extensive genetic diversity, which includes seven major genotypes and numerous subtypes [80]. Structure-based

vaccine design, leveraging insights into the E1E2 glycoprotein complex, aims to optimize antigens capable of eliciting broadly neutralizing antibodies [59]. Additionally, lipid-based nano vaccines have demonstrated enhanced neutralizing antibody responses, while polymer-based nanoparticles are being explored for their ability to stabilize antigens and improve immunogenicity, paving the way for more effective vaccines [81].

Hepatitis D virus (HDV) vaccine development presents unique challenges due to the virus's dependence on HBV for replication and its limited genome size, which restricts potential vaccine targets. Current strategies focus on experimental approaches, such as targeting HDV's farnesyltransferase pathway to inhibit replication [72]. Virus-like particles (VLPs) are also being investigated for their potential to elicit strong immune responses while overcoming the limited antigenic diversity of HDV [73]. Advancing preclinical models is critical to overcoming barriers in HCV and HDV vaccine research. The development of humanized mouse models and liver organoids provides more accurate simulations of human infection dynamics, improving the evaluation of vaccine efficacy and safety [78].

6.0.CONCLUSION

The spread of many infectious agents, including the hepatitis viruses, can be controlled by vaccination. However, there is a need to clearly understand the molecular perspectives of the design of these vaccines and they can be improved. While viruses like HAV, HBV, and HEV have approved vaccines already, HCV and HDV still lack approved vaccines because of the molecular complexities of the viruses. Future research should focus on further understanding these complexities and how they can be addressed. The development of universal hepatitis virus vaccines should also receive more attention among researchers. With continuous focus on not just the public approaches, but the molecular approaches as well, the goal of reducing the global burden of hepatitis will be achieved in the next few years.

Ethics Approval and Consent to Participate

Not applicable

Consent for Publication

Not applicable

Competing Interests

- The authors declare that they have no competing interests

Funding

- No funding was received for this study.

Authors' Contributions

- ROA conceptualized the study, drafted the manuscript, edited, proofread, and approved the final manuscript draft for submission
- AO, DOE, and SGU drafted the manuscript and approved the final draft for submission

Acknowledgments

Not applicable

Clinical Trial Number

Not applicable

HIGHLIGHTS

- While viruses like HAV, HBV, and HEV have approved vaccines already, HCV and HDV still lack approved vaccines because of the molecular complexities of the viruses.
- The development of universal hepatitis virus vaccines should also receive more attention among researchers.
- With continuous focus on not just the public approaches, but the molecular approaches as well, the goal of reducing the global burden of hepatitis will be achieved in the next few years.

REFERENCES

 Ondondo RO, Muthusi J, Oramisi V, Kimani D, Ochwoto M, Young P, et al.Prevalence of hepatitis B virus infection in Kenya: A study nested in the Kenya Population-based HIV Impact Assessment 2018. PLoS One. 2024 Nov 14;19(11):e0310923.

- 2. World Health Organization. Immunization Agenda 2030: A global strategy to leave no one behind. WHO; 2020 [cited 2025 Mar 26]. Available from: https://www.who.int/publications/m/item/imm unization-agenda-2030-a-global-strategy-to-leave-no-one-behind
- 3. Lemon SM, Ott JJ, Van Damme P, Shouval D. Type A viral hepatitis: A summary and update on the molecular virology, epidemiology, pathogenesis and prevention. *J Hepatol.* 2017 Sep 5:S0168-8278(17)32278-X.
- 4. Van Damme P, Pintó RM, Feng Z, Cui F, Gentile A, Shouval D. Hepatitis A virus infection. *Nat Rev Dis Primers*. 2023 Sep 28;9(1):51.
- 5. Iannacone M, Guidotti LG. Immunobiology and pathogenesis of hepatitis B virus infection. *Nat Rev Immunol.* 2021;22(1):19-32.
- 6. Tsukuda S, Watashi K. Hepatitis B virus biology and life cycle. *Antiviral Res*. 2020;182:104925.
- 7. Wang R, Xie Z. Non-hepatotropic viral hepatitis and its causative pathogens: The ongoing need for monitoring in children with severe acute hepatitis of unknown etiology. *Pediatr Investig.* 2022;6(3):151–155.
- 8. Yuen MF, Chen DS, Dusheiko GM, Janssen HLA, Lau DTY, Locarnini SA, Peters MG, Lai CL. Hepatitis B virus infection. *Nat Rev Dis Primers*. 2018 Jun 7;4:18035.
- 9. Urban S, Neumann-Haefelin C, Lampertico P. Hepatitis D virus in 2021: Virology, immunology and new treatment approaches for a difficult-to-treat disease. *Gut.* 2021;70(9):1782-1794. doi:10.1136/gutjnl-2020-323888.
- Le Gal F, Gault E, Ripault MP, Serpaggi J, Trinchet JC, Gordian E, Dény P. Eighth major clade for hepatitis delta virus. *Emerg Infect Dis.* 2006 Sep;12(9):1447-50.
- Georgel P, Schuster C, Zeisel MB, Stoll-Keller F, Berg T, Bahram S, Baumert TF. Virus-host interactions in hepatitis C virus infection: Implications for molecular pathogenesis and antiviral strategies. *Trends Mol Med*. 2010;16(6):277–86.
- 12. Lindenbach BD, Evans MJ, Syder AJ, Wölk B, Tellinghuisen TL, Liu CC, et al. Complete replication of hepatitis C virus in cell culture. *Science*. 2005 Jul 22;309(5734):623-6.

- 13. Tzarum N, Wilson IA, Law M. The neutralizing face of hepatitis C virus E2 envelope glycoprotein. *Front Immunol*. 2018;9:1315.
- Jopling CL, Yi M, Lancaster AM, Lemon SM, Sarnow P. Modulation of hepatitis C virus RNA abundance by a liver-specific microRNA. *Science*. 2005 Sep 2;309(5740):1577-81.
- Dreux M, Gastaminza P, Wieland SF, Chisari FV. The autophagy machinery is required to initiate hepatitis C virus replication. *Proc Natl Acad Sci U S A*. 2009 Aug 18;106(33):14046-51
- 16. Kaul A, Stauffer S, Berger C, Pertel T, Schmitt J, Kallis S, Zayas M, Lohmann V, Luban J, Bartenschlager R. Essential role of cyclophilin A for hepatitis C virus replication and virus production and possible link to polyprotein cleavage kinetics. *PLoS Pathog.* 2009 Aug;5(8):e1000546. Erratum in: *PLoS Pathog.* 2009 Sep;5(9).
- 17. Benga WJ, Krieger SE, Dimitrova M, Zeisel MB, Parnot M, Lupberger J, Hildt E, Luo G, McLauchlan J, Baumert TF, Schuster C. Apolipoprotein E interacts with hepatitis C virus nonstructural protein 5A and determines assembly of infectious particles. *Hepatology*. 2010 Jan;51(1):43-53.
- Das A, Rivera-Serrano EE, Yin X, Walker CM, Feng Z, Lemon SM. Cell entry and release of quasi-enveloped human hepatitis viruses. *Nat Rev Microbiol*. 2023;21(9):573-589.
- 19. Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. *Cell Mol Life Sci.* 2017;75(2):193-208.
- Yin X, Ambardekar C, Lu Y, Feng Z. Distinct entry mechanisms for nonenveloped and quasienveloped hepatitis E viruses. *J Virol*. 2016 Mar 28;90(8):4232-42.
- 21. Kolter T, Sandhoff K. Lysosomal degradation of membrane lipids. *FEBS Lett.* 2009;584(9):1700-12.
- 22. Surjit M, Jameel S, Lal SK. Cytoplasmic localization of the ORF2 protein of the hepatitis E virus is dependent on its ability to undergo retrotranslocation from the endoplasmic reticulum. *J Virol.* 2007;81(7):3339-45.
- 23. Elbahrawy A, Atalla H, Alboraie M, Alwassief A, Madian A, El Fayoumie M, Tabll AA, Aly

- HH. Recent advances in Protective Vaccines against Hepatitis Viruses: A Narrative Review. *Viruses*. 2023;15(1):214.
- 24. Pattyn J, Hendrickx G, Vorsters A, Van Damme P. Hepatitis B vaccines. *J Infect Dis.* 2021;224(4):S343–S351.
- Abdelwahab KS, Ahmed Said ZN. Status of hepatitis C virus vaccination: Recent update. World J Gastroenterol. 2016;22(2):862-873.
- 26. Shayeghpour A, Kianfar R, Hosseini P, Ajorloo M, Aghajanian S, Hedayat Yaghoobi M, et al. Hepatitis C virus DNA vaccines: a systematic review. *Virol J.* 2021;18(1):248.
- 27. Adugna A. Therapeutic strategies and promising vaccine for hepatitis C virus infection. Immunity, *Inflammation and Disease*. 2023;11(1):34-45.
- 28. Zahmanova G, Takova K, Tonova V, Koynarski T, Lukov LL, Minkov I et al. The re-emergence of hepatitis E virus in Europe and vaccine development. *Viruses*. 2023 Jul;15(7):1558.
- 29. Wang X, Xu W, Hu Y, Yao H, Xu L, Ma M, Zhang Y, Wang Y. Immunogenicity persistence of hepatitis A vaccines Healive® and Havrix® among children: 15 years follow-up and long-term prediction. *Hum Vaccin Immunother*. 2023;19(2):2227549.
- 30. Mahmood F, Xu R, Awan MUN, Song Y, Han Q, Xia Xet al. HBV vaccines: Advances and development. *Vaccines*. 2023;11(12):1862.
- 31. Kayesh MEH, Kohara M, Tsukiyama-Kohara K. TLR agonists as vaccine adjuvants in the prevention of viral infections: an overview. *Front Microbiol.* 2023;14:1249718.
- 32. Gomez-Escobar E, Roingeard P, Beaumont E. Current Hepatitis C vaccine candidates based on the induction of neutralizing antibodies. *Viruses.* 2023;15(5):1151.
- 33. Huang X, Wang X, Zhang J, Xia N, Zhao Q. Escherichia coli-derived virus-like particles in vaccine development. *NPJ Vaccines*. 2017;2:3.
- 34. Zuckerman A. Subunit, recombinant and synthetic hepatitis B vaccines. *Scand J Infect Dis* Suppl. 1985;47:90-5.
- 35. Francis M. Use of hepatitis B core as a vehicle for presenting viral antigens. *Rev Med Virol*. 1992;2(4):241-248.
- 36. Ondieki M, Nyaribari MC, Ejekwumadu NJ, Nyakang'o MJ. Design of a recombinant

- hepatitis B vaccine based on stably binding HLA-I peptides. *J Infect Dis Ther*. 2015;3(2):1–5.
- 37. Wu T, Li S, Zhang J, Ng M, Xia N, Zhao Q. Hepatitis E vaccine development. *Hum Vaccin Immunother*. 2012;8(6):812-23.
- Cova L. Advances and challenges in the development of therapeutic DNA vaccines against hepatitis B virus infection. *Curr Mol Med.* 2014;14(4):628-636.
- 39. Tang LL, Liu KZ. Recent advances in DNA vaccine of hepatitis virus. *Acta Pharmacol Sin.* 2002;23(5):410-8.
- 40. André F. Development and clinical studies with a recombinant DNA hepatitis B vaccine. *Vaccine*. 1989;7(2):95-8.
- 41. Zhao H, Zhou X, Zhou Y. Hepatitis B vaccine development and implementation. *Hum Vaccin Immunother*. 2020;16(3):566-576.
- 42. Davidopoulou C, Kouvelas D, Ouranidis A. Comparing vaccine manufacturing technologies: recombinant DNA vs in vitro transcribed (IVT) mRNA. Sci Rep. 2024;14(1). doi: 10.1038/s41598-024-67797-x.
- 43. Ali MZ, Anwar Y, Abbas A, Ain NU, Khalid Z, Sattar A, Usman M, Nazir A, Zeb S. Recent advances in vaccine technology for viral infection management: A spotlight on next-generation vaccines and nucleic acid-based platforms. *Pakistan Journal of Medical & Health Sciences*. 2023;17(5):261-7.
- 44. Maleksabet N, Ziaee A. Expression of S and pre-S2 hepatitis B surface antigens in mammalian Cos-7 cell line. *J Biomed Res*. 2003;1(3):135-43. Available from: https://www.ijbiotech.com/article_6900_27ef4 705d406773e225a361474a6b756.pdf
- 45. Fuyangxin, et al. Application of PreS1 in preparation of hepatitis B vaccine and treatment of chronic hepatitis B. J Hepatol. 2016. Available from: https://patentimages.storage.googleapis.com/69/b3/1f/dff20ecaa3d441/CN106421774A.pdf
- 46. Kenjiro N, Nakamura H, Kuroo M. Vaccine against hepatitis C virus and diagnostic HBC/HCV E2 peptide chimeric protein. *Immunol Vaccine Sci J.* 2015. Available at: https://patentimages.storage.googleapis.com/be/0c/cb/7b1ce8d8e1e272/WO2015133467A1.pd f

- 47. Fazlalipour M, Keyvani H, Monavari S, Mollaie H. Recombinant CoreE1E2 Protein Expressed in Pichia pastoris Yeast: A Candidate Vaccine for Hepatitis C Virus. J Vaccinol. 2014;12(3):112-120.
- 48. Dehghan Z, Lari A, Yarian F, Ahangarzadeh S, Sharifnia Z, Shahzamani K, Shahidi S. Development of polyepitopic immunogenic constructs against hepatitis C virus 1a-6a genotype by in silico approach. *Biomed Biotechnol Res J.* 2020;4(4):288-296.
- 49. Roingeard P, Hourioux C, Patient R. New protein fusion and their application for the preparation of vaccines against hepatitis C. *Vaccine Research Journal*. Available at: https://patents.google.com/patent/TW201008576A/en
- 50. Roy P, Noad R. Virus-like particles as a vaccine delivery system: Myths and facts. *Hum Vaccin*. 2008;4(1):5-12.
- 51. Mazalovska M, Kouokam JC. Progress in the production of virus-like particles for vaccination against hepatitis E virus. *Viruses*. 2020;12(8):826.
- 52. Adamowicz P, Mévélec M, Girard M. Method of preparing a vaccine against hepatitis B obtained from CHO cells. *Vaccine J.* 1986. Available from: https://patentimages.storage.googleapis.com/17/53/f3/536e6a30358e4f/CN110694059A.pdf
- 53. Prentoe J, Janitzek CM, Velázquez-Moctezuma R, Soerensen A, Jørgensen T, Clemmensen S, Soroka V, Thrane S, Theander T, Nielsen MA, Salanti A, Bukh J, Sander AF. A two-component vaccine consisting of virus-like particles displaying hepatitis C virus envelope protein 2 oligomers. *NPJ Vaccines*. 2022 Nov 15;7(1):148. Tornesello AL, Tagliamonte M, Buonaguro FM, Tornesello ML, Buonaguro L. Virus-like particles as preventive and therapeutic cancer vaccines. Vaccines (Basel). 2022 Feb 2;10(2):227.
- 54. Zuckerman A. Subunit, recombinant and synthetic hepatitis B vaccines. *Scand J Infect Dis* Suppl. 1985;47:90-5.
- 55. Pantazica AM, van Eerde A, Dobrica MO, Caras I, Ionescu I, Costache A, Tucureanu C, Steen H, Lazar C, Heldal I, Haugslien S, Onu A, Stavaru C, Branza-Nichita N, Liu Clarke J. The "humanized" N-glycosylation pathway in CRISPR/Cas9-edited Nicotiana benthamiana

- significantly enhances the immunogenicity of a S/preS1 Hepatitis B Virus antigen and the virus-neutralizing antibody response in vaccinated mice. *Plant Biotechnol J.* 2023 Jun;21(6):1176-1190. doi: 10.1111/pbi.14028.
- 56. Lawko N, Plaskasovitis C, Stokes C, Abelseth L, Fraser I, Sharma R, Kirsch R, Hasan M, Abelseth E, Willerth SM. 3D tissue models as an effective tool for studying viruses and vaccine development. Front Mater. 2021;8:631373.
- Law M. Antibody responses in hepatitis C infection. *Cold Spring* Harb Perspect Med. 2020;11(3):a036962.
- 58. Kanai T, Hu Z, Yang R, Wu W, Wang Z, D C, MA, Sanchez-Meza J, Law M, Houghton M, Law JL, Logan M, De Val N, Liang TJ. Three-dimensional reconstruction of the hepatitis C virus envelope glycoprotein E1E2 heterodimer by electron microscopic analysis. *J Virol*. 2022;97(1).
- 59. Grewal S, Hegde N, Yanow S. Integrating machine learning to advance epitope mapping. Front *Immunol.* 2024;15:1463931.
- Buschmann MD, Carrasco MJ, Alishetty S, Paige M, Alameh MG, Weissman D. Nanomaterial delivery systems for mRNA vaccines. Vaccines (Basel). 2021 Jan 19;9(1):65. doi: 10.3390/vaccines9010065.
- 61. Yoshida J. Nanopartículas de quitosana como veículo de vacinação contra a hepatite B via nasal [thesis]. São Paulo: Universidade de São Paulo; 2012. Available from: https://www.teses.usp.br/teses/disponiveis/9/91 34/tde-15032013-
 - 163603/publico/Jony_takao_yoshida_corrigida .pdf
- 62. Wang H, Ding Y, Su S, Meng D, Mujeeb A, Wu Y, Nie G. Assembly of hepatitis E vaccine by 'in situ' growth of gold clusters as nanoadjuvants: an efficient way to enhance the immune responses of vaccination. *Nanoscale Horizons*. 2016;1(5):394–398.
- 63. Zhao H, Shao X, Yu Y, Huang L, Amor NP, Guo Ket al. A therapeutic hepatitis B mRNA vaccine with strong immunogenicity and persistent virological suppression. *bioRxiv*. 2022 Nov 18.
- 64. Schmidt C, Schnierle B. Self-amplifying RNA vaccine candidates: Alternative platforms for

- mRNA vaccine development. *Pathogens*. 2023;12(1):138.
- 65. Perenkov AD, Sergeeva AD, Vedunova M, Krysko D. In vitro transcribed RNA-based platform vaccines: Past, present, and future. *Vaccines*. 2023;11(10):1600.
- 66. Roukens A, Visser L. Hepatitis B vaccination strategy in vaccine low and non-responders: A matter of quantity or quality? *Hum Vaccin*. 2011;7(6):645-648.
- 67. Buisson Y, Meyran M. Prevention of hepatitis A and hepatitis B with vaccination. *Rev Mal Respir*. 1995;179(2):367-74. Available from: https://europepmc.org/article/med/7614064
- 68. Zhu T. Study of immune evasion mechanisms of chronic hepatitis B virus in chronic hepatitis B patients. *Mod Med Sci Ess.* 2024;6(3):199-211.
- Bailey JR, Barnes E, Cox AL. Approaches, progress, and challenges to hepatitis C vaccine development. *Gastroenterology*. 2019 Feb;156(2):418-430.
- 70. Duncan J, Urbanowicz R, Tarr A, Ball J. Hepatitis C Virus Vaccine: Challenges and Prospects. *Vaccines*. 2020;8(1):90.
- Vogt A, Wohlfart S, Urban S, Mier W. Medical advances in hepatitis D therapy: Molecular targets. *Int J Mol Sci.* 2022;23(18):10817.
- 72. Collett S, Earnest L, Carrera Montoya J, Edeling MA, Yap A, et al. Development of virus-like particles with inbuilt immunostimulatory properties vaccine as candidates. Front Microbiol. 2023 Jun 7;14:1065609.
- 73. Qiu J, Zhang S, Feng Y, Su X, Cai J, Chen S et al. Efficacy and safety of hepatitis B vaccine: an umbrella review of meta-analyses. *Expert Rev Vaccines*. 2023 Jan;22(1):45-56.
- 74. De Pooter D, Pierson W, Pourshahian S, Dockx K, De Clerck B, Najera I, Davis H, Van Gulck E, Boden D. mRNA therapeutic vaccine for hepatitis B demonstrates immunogenicity and efficacy in the AAV-HBV mouse model. *Vaccines (Basel)*. 2024 Feb 25;12(3):237.
- 75. Agbede GT, Emezirinwune D, Adedokun T, Idowu-Collins P. Vaccine hesitancy in Nigeria: Overcoming cultural, linguistic, and religious obstacles. Information Impact: Journal of

DOI: 10.21608/aeji.2025.376694.1468

- Information and Knowledge Management. 2024;15(1):34-45.
- 76. Di Lello FA, Martínez AP, Flichman DM. Insights into induction of the immune response by the hepatitis B vaccine. *World J Gastroenterol.* 2022;28(31):4249-4262.
- 77. Atcheson E, Li W, Bliss CM, Chinnakannan S, Heim K, Sharpe H, et al. Use of an outbred rat Hepacivirus challenge model for design and evaluation of the efficacy of different immunization strategies for hepatitis C virus. *Hepatology*. 2020 Mar;71(3):794-807.
- 78. Costa B, Gouveia M, Vale N. Safety and efficacy of antiviral drugs and vaccines in pregnant women: Insights from physiologically based pharmacokinetic modeling and

- integration of viral infection dynamics. *Vaccines*. 2024;12(7):782.
- 79. Johnson J, Freedman H, Logan M, Wong JAJ, Hockman D, Chen C, et al. A recombinant hepatitis C virus genotype 1a E1/E2 envelope glycoprotein vaccine elicits antibodies that differentially neutralize closely related 2a strains through interactions of the N-terminal hypervariable region 1 of E2 with scavenger receptor *B1*. *J Virol*. 2019 Oct 29;93(22):e00810-19.
- 80. Bazzill J, Ochyl LJ, Giang E, Castillo S, Law M, Moon JJ. Interrogation of antigen display on individual vaccine nanoparticles for achieving neutralizing antibody responses against hepatitis C virus. *Nano Lett.* 2018 Oct;18(10):5932-5939.

Cite as: Abdullateef, R., Opatola, A., Esanju, D., Usin, S. Molecular Insights into the Development and Efficacy of Hepatitis Virus Vaccines. *Afro-Egyptian Journal of Infectious and Endemic Diseases*, 2025; (): -. doi: 10.21608/aeji.2025.376694.1468