Impact of Hepatic Steatosis on Fibrosis and Treatment Response in Chronic Hepatitis B: a Longitudinal Cohort Study

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Corresponding Author Mona Mohammed Abdelrahman Email:monamohamed@m ed sohag edu eg ORCID: 0000-0002-8784-848X. Telephone: 00201021025895 © 2024 The author (s). Published by Zagazig University. This is an open-access article under the CC BY 4.0 license https://creativecommons.o rg/licenses/bv/4.0/. Receive date: 11/7/2024 Revise date:15/10/2024 Accept date:11/11/2024 Publish date: 24/11/2024 Keywords: NAFLD, hepatic steatosis, chronic HBV, fibrosis, complete virological response.

Background and study aim: Data on the prevalence of NAFLD in chronic hepatitis B (CHB) patients is widely variable. Data about the effect of hepatic steatosis (HS) on CHB patients in the Middle East are scarce. So, we aimed to assess the associations between biopsy-proven HS and HBV viral load, liver biochemistry, and liver fibrosis and inflammation in CHB patients. We also aimed to evaluate the effect of HS on the complete virological response (CVR) within 2 years of follow-up.

Patients and Methods: This hospitalbased longitudinal cohort study included all CHB patients who were subjected to liver biopsy between June 2019 to June 2020. Patients were followed up for liver biochemistry and HBV DNA at six months' intervals for two years. CVR was defined as undetectable HBV DNA by PCR testing 27 with a detection limit of 16 IU/ml as measured by the local laboratories. The baseline clinical characteristics and accumulative incidence of CVR were compared between patients with and without HS.

INTRODUCTION

Despite the existence of an appropriate vaccination, hepatitis B virus (HBV) infection continues to be a serious health issue, with an estimated 296 million patients of chronic hepatitis В (CHB) [1]. Patients were classified according to the phase of CHB infection into: hepatitis B e antigen (HBeAg) HBV positive chronic infection. HBeAg-positive chronic hepatitis B, HBeAg-negative chronic HBV HBeAg-negative infection, and chronic hepatitis B [2]. Owing to the increased global incidence of obesity and metabolic syndrome, nonalcoholic fatty liver disease (NAFLD) has emerged as а significant **Results:** A total of 91 CHB patients were enrolled from June 2019 to June 2020. 43 (47%) of them had histological evidence of HS. Patients with chronic hepatitis had significantly higher body mass index (BMI) compared to patients with chronic infection. Patients with HS had higher HBV deoxyribonucleic acid (DNA) but without statistical significance. The severity of fibrosis was greater in HS group but without statistical significance. CVR at 6, 12, and 24 months after treatment onset was significantly higher in patients without HS (p = 0.01. 0.000,0.06, respectively).

Conclusion: HS is highly prevalent in CHB patients. HS is not associated with HBV DNA or the severity of fibrosis. The rates of CVR are higher in CHB patients without HS.

contributor to chronic liver disease. The overall prevalence of NAFLD is increasing over time at an alarming rate with an estimated worldwide prevalence of 37.8% in or after 2016 compared to 25.5% in or before 2005 [3]. As a result, the co-existence of NAFLD and HBV infection is frequently observed in healthcare settings Data on the prevalence of NAFLD in CHB patients is widely variable due to the variability of research cohorts and differences in NAFLD diagnostic techniques such as liver biopsy, transient elastography, and ultrasound. The estimated prevalence of NAFLD in CHB patients ranges from 14% to 70% [4,5].

Because both NAFLD and CHB infection share the ability to cause progression of the hepatic inflammation and fibrosis, the interaction between these two diseases has become more and more interesting to research [6,7]. However, the effect of NAFLD on the outcome of CHB infection is not well established, and various studies showed conflicting results. Most of the previous studies concluded that hepatic steatosis (HS) was associated with hepatitis B surface antigen (HBsAg) seroclearance [8-12] and low HBV deoxyribonucleic acid (DNA) serum levels [13, 14]. However, other studies found that HS did not affect HBV viral load [15,16]. HS was found to be a contributing factor in liver fibrosis progression and HCC development [12, 17-19]. On the contrary, multiple studies have revealed that HS is not a risk factor for fibrosis progression or HCC development in CHB patients [6,20, 21]. According to a recent study, CHB cases with high controlled attenuation parameter (CAP) measures had incomplete response to antiviral medications [22]. Other studies, however, revealed no connection between NAFLD and long-term biochemical or virological response to antiviral therapy [20]. To our best knowledge, data about the effect of HS on CHB patients in the Middle East are scarce; therefore, we conducted this study aiming to assess the associations between biopsy-proven HS and HBV viral load, liver biochemistry, and liver fibrosis and inflammation in CHB patients. This study also aims to evaluate the effect of HS on the complete virological response (CVR) within 2 years of follow-up.

PATIENTS AND METHODS

Study subjects and design:

This hospital-based longitudinal cohort study included all CHB patients who were subjected to liver biopsy between June 2019 to June 2020. CHB was defined as the HBsAg's continued existence for at least six months [23]. Patients were recruited from the outpatient clinic of the Medicine and Gastroenterology Tropical Department, Sohag University Hospital. The inclusion criteria were: (a) age ≥ 18 years; (b) treatment naïve; (c) detectable HBV DNA at baseline. The exclusion criteria were as follows: (a) decompensated cirrhosis; (b) HCC; (c) serological evidence of hepatitis C virus (HCV) infection; (d) human immunodeficiency virus (HIV) co-infection; (e) other causes of chronic liver diseases as autoimmune hepatitis, primary biliarv cirrhosis. Wilson's disease. hemochromatosis, or drug-induced chronic hepatitis; (f) alcohol consumption; (g) current pregnancy; (h) any contraindications to liver biopsy, such as unwilling patients, prothrombin time (PT) >4 seconds more than control; international normalized ratio (INR) greater than 1.6; platelets count <100.000/mm³. Patients were classified according to the phase of CHB infection into: hepatitis B e antigen (HBeAg) positive chronic HBV infection, HBeAg-positive chronic hepatitis B, HBeAg-negative chronic HBV infection, and HBeAg-negative chronic hepatitis B [2]. We categorized the patients into two groups: patients with chronic infection (including HBeAg positive and HBeAg-negative chronic HBV infection), and patients with chronic hepatitis (including HBeAg-positive and HBeAg-negative chronic hepatitis B).

Clinical and Laboratory Evaluation:

Baseline data included: demographic data, mellitus history of diabetes (DM)or hypertension, and body mass index (BMI) which was assessed as weight (kg) divided by height squared (m²). Liver biochemistry, HBV DNA, HBV serological markers, and a complete blood count (CBC) were also done. Abdominal ultrasound was done to evaluate liver echogenicity, and splenic size, and to exclude hepatic focal lesions.

Liver Biopsy and Histopathological Assessment:

Ultrasound-guided percutaneous liver biopsy was done using a 16G semi-automated tru-cut needle to obtain two cores of tissue, 10 mm in length each [24]. All specimens were sent to the Pathology Laboratory of the same hospital. Formalin-fixed paraffin-embedded tissue sections were prepared and processed to be stained using hematoxylin and eosin (H&E) stain. The specimens were assessed by one experienced histopathologist who is blinded to participant data. Specimens were subjected to histopathological evaluation to determine both the stage of fibrosis and the grade of inflammation. using **METAVIR** the classification system [25], which evaluates the fibrosis stage on a scale from F0 to F4. The necro-inflammatory activity was scored on a

scale from A0 to A3. A standard pathological method. including H&E, was used for examination. At least 6 portal tracts were analyzed. We classified fibrosis according to the METAVIR staging system, where F0= absence of fibrosis, F1= portal fibrosis, F2= portal and septal fibrosis, F3= nodular fibrosis, and F4= cirrhosis. Patients were divided into 2 groups according to fibrosis level: early fibrosis (mild to moderate) was defined as F0-F2; while advanced fibrosis was defined as F3-F4. NAFLD and CHB use different scoring systems for histological examination. Histologic grading of steatosis was evaluated according to the NAFLD activity score (NAS) which includes 3 components: steatosis, and lobular inflammation, ballooning degeneration. This system considers only macrovesicular steatosis and assesses the percentage of hepatocytes with steatotic vacuoles

(Figure 1). The degree of steatosis was determined as follows: Score 0 = <5%, Score 1 = 5-33%, Score 2 = 34-66%, and Score 3 = >66%. S0, S1, S2, and S3 were designated as minimal, mild, moderate, and severe steatosis, respectively. Lobular inflammation was scaled on a 0-2 scale, and ballooning on a 0-2 scale [26].

Follow UP:

Patients were followed up for liver biochemistry and HBV DNA at six months intervals for two years. Patients who were eligible to therapy treated with the available oral nucleotide analogue. CVR was defined as undetectable HBV DNA by polymerase chain reaction (PCR) testing [27] with a detection limit of 16 IU/ml as measured by the local laboratories.

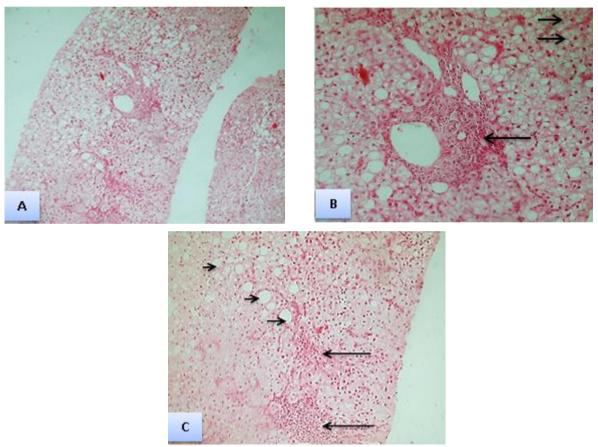


Figure 1: (A)&(B) Liver biopsy showed an area surrounding the central vein region with steatosis, inflammatory cells (long arrow), and ballooning of hepatocytes with enlarged rarefied cytoplasm (short arrow) (H&E stain, X100 & X200, respectively). (C) Hepatocytes containing large fat droplets occupying the cytoplasm and displacing the nucleus to the periphery (short arrows), foci of lobular inflammation (long arrow) composed predominantly of lymphocytes and Kupffer cells (H&E stain, X200).

Statistical analysis

SPSS version 16 (SPSS Inc., Chicago, USA) was used to assess the data. Categorical data were expressed as numbers and percentages, whereas continuous data were expressed as mean \pm standard deviation (SD) or median and interquartile range (IQR). For continuous data, examination of the differences between two groups was done using the *Student t-test* or *Mann-Whitney U* test, while the *Chi-square* test was used to analyze categorical data. The complete virological response over time was assessed using the *Kaplan-Meier* survival analysis test. *Pie and bar* charts were expressed by excel.

RESULTS

The current study was conducted on 91 CHB patients. Their mean age was 36±9.7 years with an age range of 20-60 years. Of them,84 patients (92%) were HBeAg negative and only seven (8%) patients were HBeAg positive. Sixty patients (69%) had positive hepatitis B e antibody (Anti-HBe). When evaluating the studied population according to the CHB stage [23], we found that only one patient (1%) had HBeAg positive chronic HBV infection, seven patients (8%) had HBeAg positive chronic hepatitis, 29 patients (32%) had HBeAg negative chronic HBV infection, and 54 patients (59%) had HBeAg negative chronic hepatitis (Figure1). Patients with chronic hepatitis had significantly higher BMI and alanine transaminase (ALT) levels compared to patients with chronic infection (p = 0.003, 0.01, respectively). The prevalence of HS was higher in patients with chronic hepatitis compared to patients with chronic infection but without statistical significance. Patients with chronic hepatitis had significantly higher scores of lobular inflammations compared to those with chronic infection (p=0.02) (Table 1). Forty-seven percent of the studied cohort (43 patients) showed HS on liver biopsy (Figure 2). A mild degree of HS was reported in 28 CHB patients (31%), a minimal degree of HS was reported in 11 patients (12%), and a moderate degree of HS was reported in 4 CHB patients (4%). However, a severe degree of HS was not reported (Figure 3). According to the presence of steatosis on liver biopsy, we divided our population into two groups: the first group included CHB patients with HS (n=43), while the second group included CHB patients without HS (n=48). The age and gender distribution had no significant difference between the two groups. The BMI, aspartate transaminase (AST), total bilirubin, and splenic size were significantly higher in CHB patients with HS than those without (p = 0.02, 0.03, 0.001, respectively). Moreover, patients with HS had higher HBV DNA and ALT levels but without statistical significance. The severity of hepatic fibrosis was greater in patients with HS than those without steatosis but without a statistical significance (p = 0.08) (Table 2). High lobular inflammation scores had a strong significant association with advanced fibrosis stages (P=0.000) (Table3).

All patients with HBeAg positive and negative hepatitis (61 patients) received medical treatment in the form of lamivudine 100 mg tablets once daily. During follow up six patients were excluded due to poor compliance. Fiftyfive patients were followed for 2 years to evaluate the virological response. CVR at 6 months, 12 months, and 24 months after treatment onset was significantly higher in CHB patients without HS compared to those with HS (p = 0.01, 0.000, 0.06, respectively). As CVR at 6th month of treatment was reported in 88% of CHB patients without HS versus 57% of those with HS. Moreover, after 1 years of follow up 100% of CHB without HS achieved CVR, while only 56.7% of those with HS achieved CVR with one patient reported virological breakthrough (Table 4).

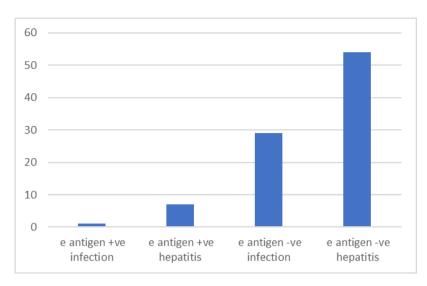


Figure 2: Stages of CHB infection in the studied population (Bar chart).

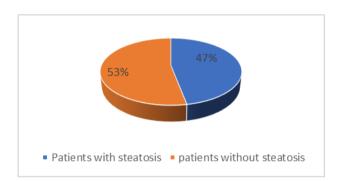


Figure 3: Percentage of CHB patients with steatosis on liver biopsy (Pie chart).

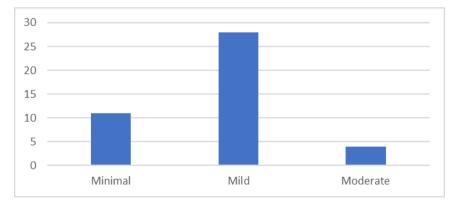


Figure 4: Different degrees of steatosis in the studied population (Bar chart).

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Table 1: Baseline clinical, laboratory and histopathological characteristics of the studied population according to CHB stage (N=91).

	Chronic infection*		
	N=30	Chronic hepatitis** N=61	P-value
Age (years), mean±SD	36±11.2	35.7±9	0.6
BMI, median (range)	22.7	25.7	0.003
	(20.5-30)	(23.5-30)	
Male, n%	24 (80%)	45 (73.8%)	0.5
Female, n%	6 (20%)	16(26.2%)	
HBV DNA (IU/L), median	8700	15000	0.2
(IQR)	(5324-53469)	(4530-91395)	
ALT (IU/L),	21.5	29	0.01
median (IQR)	(17.3-27)	(20-46.5)	
AST(IU/L),	17.8	24.6	0.3
median (IQR)	(22.5-28)	(18-38)	
Albumin(g/dl), mean±SD	4.3±0.4	4.3±0.5	0.2
Total bilirubin (mg/dl), mean±SD	0.72±0.27	0.74±0.22	0.6
PT, mean±SD	12.9±0.8	12.7±1	0.5
Prothrombin concentration, mean±SD	88.9±17.5	92±11.6	0.2
INR, mean±SD	1.05±0.12	1.04±0. 1	0.3
Steatosis	12 (40%)	31 (50.8%)	0.3
Fibrosis			0.000
F0	9(30%)	0 (0%)	

F1	21 (70 %)	15 (24.6 %)	
F2	0(0%)	35 (57.4%)	
F3	0(0%)	6(9.8%)	
F4	0 (0%)	5(8.2 %)	
Mild to moderate fibrosis (F0, F1, F2)	30 (100%)	50 (82%)	0.000
Advanced fibrosis (F3, F4)	0(0%)	11(18%)	
Cirrhosis	0(0%)	5(8.2 %)	0.01
Inflammation			0.000
A0	3(10%)	0(0%)	
A1	24 (80%)	12(19.6%)	
A2	2(6.7%)	40(65.6%)	
A3	1(3.3%)	9(14.8%)	
Lobular inflammation			0.02
0	12 (40%)	8 (13.1%)	
1	12 (40%)	26 (42.6%)	
2	5 (16.7%)	17 (27.9%)	
3	1 (3.3%)	10 (16.4%)	
Hepatocyte ballooning			0.5
0	15 (50.0%)	24 (39.3%)	
1	11 (36.7%)	24 (39.3%)	

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2	14 (13.3%)	13 (31.3%)	

Chi-square test was used to analyze categorical data

Student t-test or Mann-Whitney U test to examine of the differences between two groups

*(1 patient had HBeAg positive infection and 29 patients had HBeAg negative infection)

**(7 patients had HBeAg positive hepatitis and 54 patients HBeAg negative hepatitis)

ALT: alanine transaminase, AST: aspartate transaminase, HBV: hepatitis B virus, DNA: deoxyribonucleic acid, INR: international randomized ratio, IQR: interquartile range, BMI: body mass index, PT: prothrombin time.

Normal range of laboratory data: HBV DNA less than 20IU/L, ALT (0-40 IU /L), AST (0-40 IU /L), Albumin (3.5-5.2), T.bilirubin (0.2-1 mg/dl), PT (11-12.5), INR less than 1.1

Table 2: Baseline clinical, laboratory, and histopathological characteristics of the studied population according to the presence of steatosis (N=91).

	CHB patients with HS N=43	CHB patients without HS N=48	P-value
Age (years), mean±SD	36±8	36±10	0.9
BMI, median (IQR)	26.9	23.6	0.02
	(23.8-30)	(21.3-27)	
Male,n%	29 (67%)	40 (83%)	0.08
Female, n%	14 (33%)	8(17%)	
DM	3 (7%)	6(13%)	0.4
Hypertension	2 (5%)	1(2%)	0.4
Abdominal ultrasound measurements, mean±SD	13.8 ±1.5	14±1.6	0.4
Liver size (cm)	11.4±1.8	14 ± 1.0 10.3±1.5	0.001
Splenic size (cm)			
Hepatic echo-pattern	23(53.5%)	32(66.7%)	0.02
Normal	9(20.1%)	0(0%)	
Bright			

Γ			
Coarse	11(25.6%)	16 (33.3%)	
HBV DNA (IU/L),	17137	9500	0.2
median (IQR)	(5880- 70692)	(3639-65872.5)	
ALT (IU/L),	29	22.1	0.07
Median (IQR)	(20-44)	(16.5-48.5)	
AST(IU/L),	27	21.3	0.03
Median (IQR)	(20-33)	(16-27.5)	
Albumin(g/dl), mean±SD	4.2±0.58	4.3±0.34	0.5
Total bilirubin (mg/dl),	0.79±0.23	0.68±0.23	0.03
mean±SD			
PT, mean±SD	12.8±1	12.8±0.98	0.9
Prothrombin concentration, mean±SD	92±16.4	90±11	0.7
INR, mean±SD	1.03±0.09	1.05±0.12	0.3
Fibrosis			0.08
F0	0 (0%)	5 (10.4 %)	
F1	19 (44.2%)	15 (31.2%)	
F2	17 (39.5%)	24(50 %)	
F3	3 (7%)	3 (6.2 %)	
F4	4 (9.3%)	1(2.1 %)	
Mild to moderate fibrosis (F0, F1, F2)	36 (83.7%)	44 (91.6%)	0.2
Advanced fibrosis (F3, F4)	7(16.3%)	4(0.4%)	

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Cirrhosis	4 (9.3%)	1 (2%)	0.1
Inflammation			0.8
A0	2(4.7%)	2(4.2%)	
A1	16(37.2%)	19(39.6%)	
A2	19(44.2%)	23(47.9%)	
A3	6(14%)	4(8.3%)	

Chi-square test was used to analyze categorical data *Student t-test* or *Mann-Whitney U* test to examine of the differences between two groups

ALT: alanine transaminase, AST: aspartate transaminase, HBV: hepatitis B virus, DM: diabetes mellitus, DNA: deoxyribonucleic acid, INR: international normalized ratio, IQR: interquartile range, BMI: body mass index, PT: prothrombin time.

Normal range of laboratory data: HBV DNA less than 20IU/L, ALT (0-40 IU /L), AST (0-40 IU /L), Albumin (3.5-5.2), T.bilirubin (0.2-1 mg/dl), PT (11-12.5), INR less than 1.1

Table 3: Relationship between inflammation and ballooning and the stage of fibrosis in the

studied population (N=91)

Stage of fibrosis	Lobular inflammation			р-	Ballooning of hepatocyte			p-	
11010515	0	1	2	3	value	0	1	2	value
F0 N=5	4 (80.0%)	1 (20.0%)	0 (0.0%)	0 (0.0%)	0.000	5 (100.0%)	0 (0.0%)	0 (0.0%)	0.1
F1 N=34	8 (23.5%)	19 (55.9%)	5 (14.7%)	2 (5.9%)		13 (38.2%)	12 (35.3%)	9 (26.5%)	
F2 N=41	7 (17.1%)	18 (43.9%)	13 (31.7%)	3 (7.3%)		19 (46.3%)	17 (41.5%)	5 (12.2%)	
F3 N=6	1 (16.7%)	0 (0.0%)	2 (33.3%)	3 (50.0%)		1 (16.7%)	4 (66.7%)	1 (16.7%)	
F4 N=5	0 (0.0%)	0 (0.0%)	2 (40.0%)	3 (60.0%)		1 (20.0%)	2 (40.0%)	2 (40.0%)	

Chi-square test was used to analyze categorical data

	CHB patients with HS N= 30	CHB patients without HS N=25	P-value
CVR at 6 months, N (%)	17 (56.7%)	22 (88%)	0.01
CVR at 12 months, N (%)	16 (53%)	25(100%)	0.000
CVR at 24 months, N (%)	26 (68.7%)	25 (100%)	0.06

Table 4: The accumulative incidence of virological response at 6 months, 1 year, and 2 years after the start of therapy (N=55).

Kaplan-Meier survival analysis test

CVR: Complete virological response, HS: hepatic steatosis.

DISCUSSION

The increasing global prevalence of NAFLD raised the interest of researchers to study the interaction between NAFLD and other causes of chronic liver diseases. Our biopsy-based longitudinal cohort study evaluated different aspects of the relationship between NAFLD and CHB infection, including HBV viral load, liver biochemistry, liver fibrosis, inflammation, and the effect of HS on virological response to antiviral therapy. We found a high prevalence of biopsy-proven HS (about 50%) in CHB patients. This is consistent with previously published data from Turkey [28,29], Italy [30], Taiwan [31], and China [32]. A low prevalence of HS in CHB patients was found by other investigators (5% to 34%) [33-35]. However, these studies were based on imaging modalities rather than liver biopsy, which is the gold standard for the diagnosis and grading of HS. The effect of HBV on lipid metabolism is complex. Animal studies showed that both lipid degradation and lipid synthesis were enhanced by HBV infection [36]. HBV X protein was found to be a key factor in regulating HS. HBV X protein enhances the gene expression of different lipogenic enzymes leading to fat deposition in hepatocytes and

consequently HS [37-39]. Moreover, HBV DNA was found to be a contributing factor to HS by interfering with Sterol regulatory element binding proteins-1c (SREBP-1c) expression [40]. However, host factors such as metabolic syndrome have a predominant role in the induction of HS and cannot be neglected. In our study, we found that CHB patients with HS had a higher BMI, which is in agreement with many authors [28,31,33,41]. In addition, we found that CHB patients with active disease had a significantly higher BMI compared to patients with stable disease, while no significant difference was found between the two groups as regards HS frequency. This finding confirms the proposed role of obesity in the progression of Adiponectin, a powerful adiposefibrosis. anti-inflammatory mediator. derived is frequently deficient in obesity. Adiponectin deficiency contributes to hepatic stellate cell (HSC) activation, fibrosis, and increased production of inflammatory mediators. Animal models showed that adiponectin knockout mice kept on a high-fat diet had more severe fibrosis than wild controls [42], whereas carbon tetrachloride 4 induced fibrosis is reduced in mice after receiving adiponectin [43].

Our data showed that CHB patients with HS tended to have higher HBV DNA levels than those without HS, but without statistical significance. Previous literature investigated the relationship between HBV DNA and HS with conflicting results. Two retrospective studies from China showed that HS was associated with metabolic factors rather than viral factors; namely HBV DNA and HBeAg status [44,45]. Moreover, many other studies did not find any relation between HBV DNA levels and NAFLD in CHB patients [33,46,47]. On the other hand, Xiaoyan et al [48]. found that HBV DNA levels were significantly higher in lean CHB patients with minimal HS compared to those without HS. However, the proposed protective effect of HS in CHB patients was attributed to the inverse relationship between HS and HBV DNA documented by many authors [28,49,50]. Nevertheless, the diagnosis of HS in many of imaging-based; these studies was using abdominal ultrasound or CAP measurements which is less accurate than liver biopsy; and this is a major strength in our study. Furthermore, the mechanism of the inverse relationship between HS and HBV DNA is still unclear. To our knowledge, few studies investigated this possible mechanism, and it was claimed that HS suppresses HBsAg and HBV DNA secretion through induction directly of hepatocyte endoplasmic reticulum (ER) stress [51], or indirectly through induction of hepatocyte apoptosis ¹⁰. Undoubtedly, more thorough research is required to uncover the relationship between HS and HBV DNA and its possible mechanisms.

The current study found that the frequency of advanced fibrosis (F3 and F4 METAVIR score) was higher in HS group but without a statistical significance. HS has been identified by many authors as a factor contributing to the burden of liver fibrosis in CHB patients. Two crosssectional studies from China reported that severe steatosis measured by CAP score was associated with severe fibrosis in both treatment naïve CHB patients and those who received treatment [14,19]. A possible explanation of the association of HS with the progression of fibrosis is NAFLD-related lipotoxicity, with the production of excess free radicals through endoplasmic reticulum stress [52]. On the contrary, many authors did not find a relation between HS and the stage of fibrosis [4,53-55]. In addition, Bondini et al [56]. reported that liver fibrosis in CHB patients was related to viral and metabolic factors rather than HS. Moreover, we found that the severity of fibrosis was linked to biopsyproven lobular inflammation rather than the presence of steatosis itself, and this is partially consistent with the results of Huang et al [57] who reported a significant association of the of fibrosis with both severity lobular inflammation and cytological ballooning rather than steatosis. An interesting finding was documented by a retrospective biopsy-based study from China, as they found that HS had an inverse relationship to the grade of inflammation and the stage of fibrosis [44]. Thus, we can say that convincing evidence linking HS to hepatic fibrosis is still lacking.

Another major finding of the current study is that HS had a negative impact on the virological response to oral antiviral therapy in CHB patients. We observed significantly higher rates of CVR in the non-steatotic group at 6 months, 12 months, and 24 months after the onset of lamivudine therapy. The negative impact of HS on the treatment response was also observed by Zhu et al [41]. They found that HBeAg positive patients with HS had a lower total virological response rate at 12 weeks of entecavir therapy compared to those without HS. This is consistent with Chen et al [57]. who observed a poor response to antiviral therapy in CHB patients with high CAP measurements at 12, 24, and 48 weeks of therapy compared to those with normal CAP measurements. Another prospective study observed that the rate of HBV-DNA clearance was significantly higher at 24, 48, and 96 weeks of entecavir therapy in CHB patients without HS [58]. On the contrary, Li et al [33]. observed comparable cumulative rates of complete viral suppression in CHB NAFLD and CHB non-NAFLD groups at 12, 24, 36, 48, and 60 months of oral antiviral therapy. Despite the advantage of long follow-up duration, the study of Li et al.

used radiological studies as the main methods for diagnosis of HS, and only a few cases underwent liver biopsy. Moreover, DNA quantification was done using PCR testing with a relatively high detection limit (20-100 IU/ml) compared to that of the testing used in the current study (16 IU/ml), making the definition of CVR variable in the two studies. The possible negative effect of HS on treatment response could be attributed to hepatocellular fat droplet accumulation, which reduces the bioavailability of nucleoside analogues' intrahepatic metabolites [59]. The effectiveness of therapy may also be impacted by decreased hepatic cytochrome activity in steatotic hepatocytes, insulin resistance, and obesity coexisting with HS, which results in dysfunction of cellular immunity [58].

Despite our best efforts, this study has some limitations. First, it is a single-center study with a small sample size. However, it was difficult to readily increase the sample size due to the biopsy-based nature of the study; which would be challenging to approach in future prospective studies given the emergence of noninvasive techniques. Undoubtedly, we performed a liver biopsy only when it was indicated. Secondly, a small proportion of our cohort had positive HbeAg. Thirdly, we evaluated the virological response to lamivudine only, which was the only available oral antiviral drug in our locality at the time of data collection. Lastly, we followed our cohort for 24 months only after the start of therapy, which is a relatively short period to detect treatment resistance.

To conclude, the results of this longitudinal cohort biopsy-based study of CHB patients confirmed the high prevalence of HS among CHB patients. We also detected that CHB activity is related to BMI rather than HS. We did not find a significant association between HS with HBV DNA and the severity of fibrosis. The rates of CVR were higher in the non-HS group, emphasizing the need for further studies with large sample sizes to confirm the results and to identify if combined CHB-HS necessitate any special considerations when choosing the suitable CHB treatment line.

Ethical statement: This study conformed to the ethical guidelines of the Declaration of Helsinki and was approved by the Scientific Research Ethical Committee, Faculty of Medicine, Sohag University (IRB Registration number: Soh-Med-22-12-26).

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Authors' contribution: Conceptualization, formal analysis, and original draft writing; Mona Mohammed Abdelrahman. Resources provision, data curation management, and supervision; Amira Maher, Nagwa Abd Elsadek Ahmed. Methodology, review, and editing; all authors.

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Clinical trial registration: NCT05678582, retrospectively registered, https://clinicaltrials.gov/study/NCT05678582.

Abbreviations: ALT: alanine transaminase; AST: aspartate transaminase; BMI: body mass index; CAP: controlled attenuation parameter; CHB: chronic hepatitis B; CVR: complete virological response; DNA: deoxyribonucleic acid; HBV: hepatitis B virus; HBsAg: hepatitis B surface antigen; HCC: hepatocellular carcinoma; HS: hepatic steatosis; INR: international randomized ratio; NAFLD: non-alcoholic fatty liver disease; PT: prothrombin time.

HIGHLIGHTS:

• Because both NAFLD and CHB infection share the ability to cause progression of the hepatic inflammation and fibrosis, the

interaction between these two diseases has become extremely interesting to research.

- There was high incidence of hepatic steatosis among patients with CHB.
- Patients with chronic hepatitis had significantly higher BMI compared to patients with chronic infection.
- The rates of CVR are higher in CHB patients without HS.

REFERENCES

- 1. World Health organization. Global progress report on HIV, viral hepatitis and sexually transmitted infections, 2021: accountability for the global health sector strategies 2016– 2021: actions. Accessed January 22, 2023. https://apps.who.int/iris/bitstream/handle/106 65/342813/9789240030992-eng.pdf.
- Lampertico P, Agarwal K, Berg T, Buti M, Janssen H, Papatheodoridis G, et al. EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection. J Hepatol. 2017; 67(2):370-98.
- Riazi K, Azhari H, Charette J, Underwood F, King J, Afshar E, et al. The prevalence and incidence of NAFLD worldwide: a systematic review and meta-analysis *.Lancet Gastroenterol Hepatol.* 2022; 7(9):851-61.
- Machado MV, Oliveira, Cortez-Pinto H. Hepatic steatosis in hepatitis B virus infected patients: meta-analysis of risk factors and comparison with hepatitis C infected patients *J Gastroenterol Hepatol.* 2011; 26(9):1361-7.
- Choi HSJ, Brouwer WP, Zanjir WMR, de Man RA, Feld JJ, et al. Nonalcoholic Steatohepatitis Is Associated With Liver-Related Outcomes and All-Cause Mortality in Chronic Hepatitis B. *Hepatol.* 2020; 71(2):539-48.
- Lee YB, Ha Y, Chon YE, Kim MN, Lee JH, Park H, et al. Association between hepatic steatosis and the development of hepatocellular carcinoma in patients with chronic hepatitis B. *Clin Mol Hepatol* .2019; 25(1):52-64.
- Chan AWH, Wong GLH, Chan HY, Tong JHM, Yu Y-H, Choi PCL, et al. Concurrent fatty liver increases risk of hepatocellular carcinoma among patients with chronic hepatitis B *J Gastroenterol Hepatol*.2017; (3) ^{rr}: 667-76.

- Tai DI, Lin SM, Sheen IS, Ch C-M, Lin D-Y, Liaw Y-F. Long-term outcome of hepatitis B e antigen–negative hepatitis B surface antigen carriers in relation to changes of alanine aminotransferase levels over time. *Hepatol.* 2009; (6)49: 1859-67.
- Li J, Yang H, Yeh M, Le M, Le A, Yeo Y, et al. Association between fatty liver and cirrhosis, hepatocellular carcinoma, and hepatitis B surface antigen seroclearance in chronic hepatitis B. J Infect Dis. 2021; 224(2):294-302.
- 10. Chu CM, Lin DY, Liaw YF. Does increased body mass index with hepatic steatosis contribute to seroclearance of hepatitis B virus (HBV) surface antigen in chronic HBV infection? *Int J Obes*. 2007; 31(5):871-5.
- 11. Chu CM, Lin DY, Liaw YF. Clinical and virological characteristics post HBsAg seroclearance in hepatitis B virus carriers with hepatic steatosis versus those without. *Dig Dis Sci.* 2013; 58(1):275-81.
- Mak LY, Hui RWH, Fung J, Liu F, Wong D, Cheung K-S, et al. Diverse effects of hepatic steatosis on fibrosis progression and functional cure in virologically quiescent chronic hepatitis B. *J Hepatol.* 2020; 73(4):800-6.
- Wang L, Wang Y, Liu S, Zhai X, Zhou G, Lu F, et al. Nonalcoholic fatty liver disease is associated with lower hepatitis B viral load and antiviral response in pediatric population. *J Gastroenterol.* 2019; 54(12):1096-105.
- 14. Hui RHW, Seto W-K, Cheung K-S, Mak L-Y, Liu K, Fung J, et al. Inverse relationship between hepatic steatosis and hepatitis B viremia: results of a large case-control study . *J Viral Hepat.* 2018; 25(1):97-104.
- 15. Tsochatzis E, Papatheodoridis GV, Manesis EK, Kafiri G, Tiniakos D, Archimandritis A. Metabolic syndrome is associated with severe fibrosis in chronic viral hepatitis and non-alcoholic steatohepatitis. *Aliment Pharmacol Ther.* 2008; 27(1):80-9.
- 16. Minakari M, Molaei M, Shalmani HM, Mohammad A, Davarpanah A, Nosratollah N, et al. (2009) Liver steatosis in patients with chronic hepatitis B infection: host and viral risk factors. *Eur J Gastroenterol Hepatol*. 2009; 21(5):512-6.
- Charatcharoenwitthaya P, Pongpaibul A, Kaosombatwattana U, Bhanthumkomol P, Bandidniyamanon W, Pausawasdi N, et al. (2017). The prevalence of steatohepatitis in chronic hepatitis B patients and its impact on

disease severity and treatment response. Liver Int: $(\xi)^{\text{rv}}$. 2017; 27(4); 542-51.

- Fan R, Niu J, Ma H, Xie Q, Cheng J, Rao H, et al. (2021) Association of central obesity with hepatocellular carcinoma in patients with chronic hepatitis B receiving antiviral therapy. *Aliment Pharmacol Ther.* 2021; 54(3):329-38.
- Seto W-K, Hui R, Mak L-Y, Fung J, Cheung K-S, Liu K, et al. Association Between Hepatic Steatosis, Measured by Controlled Attenuation Parameter, and Fibrosis Burden in Chronic Hepatitis B. *Clin Gastroenterol Hepatol.* 2018; 16(4):575-83.
- Lim C, Goh G, Li H, Lim T, Leow W, Wan W, et al. Presence of hepatic steatosis does not increase the risk of hepatocellular carcinoma in patients with chronic hepatitis B over long follow-up. *Microbiol Insights*. 2020; 13:117863612091887.
- Chang JW, Lee JS, Lee HW, Kim B, Park J, Kim D, et al. No influence of hepatic steatosis on the 3-year outcomes of patients with quiescent chronic hepatitis B *.J Viral Hepat.* 2021; 28(11): 1545–53.
- Tong X, Song Y, Yin S, Wang J, Huang R, Wu C, et al. Clinical impact and mechanisms of hepatitis B virus infection concurrent with non-alcoholic fatty liver disease. *Chinese Med J* .2022;134(14):1653-63.
- Terrault NA, Lok ASF, Mcmahon BJ, Chang K-M, Hwang J, Jonas MM, et al. Update on Prevention, Diagnosis, and Treatment of Chronic Hepatitis B: AASLD 2018 Hepatitis B Guidance. *Hepatol*.2018; 67(4):1560-99.
- 24. Rockey DC, Caldwell SH, Goodman ZD Nelson RC, Smith AD. Liver biopsy. *Hepatol.* 2009; 49(3):1017-44.
- 25. Bedossa P, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. *Hepatol.* 1996; 24(2):289-93.
- 26. Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatol.* 2005; 41(6):1313-21.
- Korean Association for the Study of the Liver. KASL Clinical Practice Guidelines: Management of chronic hepatitis B. *Clin Mol Hepatol.* 2012; 18 (2):109-62.

- 28. Ceylan B, Arslan F, Batırel A, Muzaffer F, Cem Y, Esra F, et al. Impact of fatty liver on hepatitis B virus replication and virologic response to tenofovir and entecavir *.Turkish J Gastroenterol.* 2016; 27:42-8.
- 29. Sayar M, Bulut D, Acar A. Evaluation of hepatosteatosis in patients with chronic hepatitis B virus infection. Arab Journal of Gastroenterol. 2023; 24(1):11-15.
- 30. Petta S, Cammà C, Marco VD, Macaluso FS, Maida M, Pizzolanti G, et al. Hepatic steatosis and insulin resistance are associated with severe fibrosis in patients with chronic hepatitis caused by HBV or HCV infection. *Liver International.* 2011; 31(4):507-15.
- 31. Chen Y-C, Jeng W-J, Hsu C-W, Lin C-Y. Impact of hepatic steatosis on treatment response in nuclesos(t)ide analogue-treated HBeAg-positive chronic hepatitis B: A retrospective study. *BMC Gastroenterol.* 2020; 20(1):1-9.
- 32. Zheng R, Xu C, Jiang L Dou AX, Zhou K, Lu LG (2010) Predictors of hepatic steatosis in HBeAg-negative chronic hepatitis B patients and their diagnostic values in hepatic fibrosis *International journal of medical sciences*.2010; 7 (5)272-7.
- 33. Li J, Le AK, Chaung KT, Henry L, Hoang J, Cheung R, et al. Fatty liver is not independently associated with the rates of complete response to oral antiviral therapy in chronic hepatitis B patients. *Liver International.* 2020; 40(5):1052-61.
- 34. Lin YC, Hsiao ST, Chen JD. Sonographic fatty liver and hepatitis B virus carrier status: synergistic effect on liver damage in Taiwanese adults. *World J Gastroenterol.* 2007; 13(12):1805-10.
- 35. Huang, J, Jing M, Wang, C, Wang M, You S, Lin S, Zhu Y. The impact of hepatitis B virus infection status on the prevalence of nonalcoholic fatty liver disease: A population-based study. *J Med Virol.* 2020; 92: 1191–7.
- 36. Zhang J, Ling N, Lei Y, Peng M, Hu P, Chen M. Multifaceted Interaction Between Hepatitis B Virus Infection and Lipid Metabolism in Hepatocytes: A Potential Target of Antiviral Therapy for Chronic Hepatitis B. Front Microbiol. 2021; 12:636897
- 37. Kim J, Song E, Lee H, Oh Y, Choi K, Yu D, Park S, et al. HBx-induced hepatic steatosis and apoptosis are regulated by TNFR1-and

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NF-κB-dependent pathways. *Journal of molecular biology*. 2020; 397(4), 917-31.

- Kim K, Shin H, Kim K, Choi H, Rhee S, Moon H, et al. Hepatitis B virus X protein induces hepatic steatosis via transcriptional activation of SREBP1 and PPARγ. *Gastroenterology*. 2007; 132(5), 1955-67.
- 39. Lin X, Wu YL, Peng XE, Yan X, Chen W, Lin X. Hepatitis B virus X protein induces hepatic steatosis by enhancing the expression of liver fatty acid binding protein. *Journal of Virology*. 2016; 90(4), 1729-40.
- 40. Jiang CY, Zeng WQ, Chen YX, Dai FH, Jiang P. Effect of HBV on the expression of SREBP in the hepatocyte of chronic hepatitis B patients combined with hepatic fatty change. *Chinese Journal of Hepatology*. 2011 19(8):608-613.
- 41. Zhu LY, Wang YG, Wei LQ, Zhu J, Dai W, Zhang X. The effects of the insulin resistance index on the virologic response to entecavir in patients with HBeAg-positive chronic hepatitis B and nonalcoholic fatty liver disease. *Drug Design, Development and Therapy*. 2016; 10:2739-44.
- 42. Asano T, Watanabe K, Kubota N, Gunji T, Omata M, Kadowaki T, Adiponectin knockout mice on high fat diet develop fibrosing steatohepatitis. *J Gastroenterol Hepatol.* 2009 ;24(10):1669-76.
- Kamada Y, Tamura S, Kiso S, Matsumoto H, Saji Y, Yoshida Y, et al. Enhanced Carbon Tetrachloride-Induced Liver Fibrosis in Mice Lacking Adiponectin. *Gastroenterol.* 2003; 125(6):1796-807.
- 44. Shi JP, Fan JG, Wu R, Gao X-q, Zhang L, Wang H, et al. Prevalence and risk factors of hepatic steatosis and its impact on liver injury in Chinese patients with chronic hepatitis B infection. J Gastroenterol Hepatol. 2008; 23(9):1419-25.
- 45. Huang Y, Gan Q, Lai R, Wang W, Guo S, Sheng Z, et al. Application of Fatty Liver Inhibition of Progression Algorithm and Steatosis, Activity, and Fibrosis Score to Assess the Impact of Non-Alcoholic Fatty Liver on Untreated Chronic Hepatitis B Patients. *Front Cell Infect Microbiol.* 2022;11.
- 46. Tang Y, Fan R, Lan Z, Xie Q, Zhang J, Liang X, et al. Impact of nonalcoholic fatty liver disease status change on antiviral efficacy of nucleos(t)ide analogues in HBeAg-positive chronic hepatitis B. *J Med Virol.* 2023; 95(2): e28501.

- 47. Sayar MS, Bulut D, Acar A. Evaluation of hepatosteatosis in patients with chronic hepatitis B virus infection. *Arab Journal of Gastroenterology*. 2023; 24(1):11-5.
- MA X, CHEN Y, Liu J, Li J, Wu C. Clinical features and related risk factors of chronic hepatitis B patients with concomitant minimal hepatic steatosis. *J Clin Hepatol.* 2023; 39(1): 63-9.
- Leow Y-W, Chan W-K, Goh G, Wong V, Fan J, Kim Y, et al. Hepatic steatosis and metabolic risk factors among patients with chronic hepatitis B: The multicentre, prospective CAP-Asia study. *J Viral Hepat.* 2023; 30: 319- 326.
- 50. Hui RWH, Seto WK, Cheung KS, Mak L-Y, Liu KSH, Fung J, et al. Inverse relationship between hepatic steatosis and hepatitis B viremia: results of a large case-control study. *J Viral Hepat.* 2018; 25(1):97-104.
- 51. Liu Q, Mu M, Chen H, Zhang G, Yang Y, Chu J, Li Y, et al. Hepatocyte steatosis inhibits hepatitis B virus secretion via induction of endoplasmic reticulum stress. *Mol Cell Biochem.* 2022; 477(11):2481-91.
- 52. Unger R, Clark G, Scherer P, Orci L. Lipid homeostasis, lipotoxicity and the metabolic syndrome *.Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids.* 2010; 1801(3), 209-14.
- 53. Mao X, Shing Cheung K, Peng C, Mak L-Y, Cheng H, Fung J, et al. Steatosis, HBV-related HCC, cirrhosis, and HBsAg seroclearance: A systematic review and meta-analysis. *Heptology*. 2022; 00:1-11.
- 54. Thomopoulos KC, Arvaniti VT, samantas AC, Dimitra D, Charalambos G, Dimitrios S, et al. Prevalence of liver steatosis in patients with chronic hepatitis B: a study of associated factors and of relationship with fibrosis *.European journal of gastroenterology & hepatology.* 2006; 18(3), 233-7.
- 55. Zheng O, Zou B, Wu Y, Yeo W, Wu H, Stave C, et al. Systematic review with meta-analysis: prevalence of hepatic steatosis, fibrosis and associated factors in chronic hepatitis Β. Alimentary Pharmacology & Therapeutics. 2021; 54(9):1100-9.
- Bondini S, Kallman J, Wheeler A, Parkash S, Gramlich T, Jondle D, et al. Impact of non-alcoholic fatty liver disease on chronic hepatitis B. *Liver international*. 2007; 27(5):607-611.

- 57. Chen J, Wang ML, Long Q, Bai L, Tang H High value of controlled attenuation parameter predicts a poor antiviral response in patients with chronic hepatitis B. *Hepatobiliary & Pancreatic Diseases International.* 2017; 16(4):370-374.
- 58. Jin X, Chen YP, Yang YD, Li Y-M, Zheng L, Xu C. Association between hepatic steatosis and entecavir treatment failure in

chinese patients with chronic hepatitis B. *PLoS One*. 2012; 7(3): e34198.

 Lau E, Carvalho D, Freitas. Gut microbiota: association with NAFLD and metabolic disturbances. *BioMed research int.* 2015:979515. doi: 10.1155/2015/979515.

Site as: Mohammed Abdelrahman, M., Abd El-Sadek Ahmed, N., Maher, A. Impact of Hepatic Steatosis on Fibrosis and Treatment Response in Chronic Hepatitis B: a Longitudinal Cohort Study. *Afro-Egyptian Journal of Infectious and Endemic Diseases*, 2024;14(4): 487-503. doi: 10.21608/aeji.2024.303453.1398